

The Research Progress of Armillaria Mellea Polysaccharide

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Keywords: Armillaria mellea, polysaccharide, bioactive, antioxidant, antitumor

Abstract: Armillaria mellea is also known as honey mushroom ring, wheel Ye Ji mushrooms or hazel mushroom, is one of the traditional medicine combination food fungi, its polysaccharide has many kinds of biological activity, such as antioxidation, anti-inflammatory, anti-tumor and so on. With the deepening of the research, more and more honey fungus polysaccharide was found, and the study of its biological activity has also made significant progress. This paper mainly discusses the armillaria polysaccharide extraction process and the optimization, structure, and physical and chemical properties, biological activity and so on, so as to provide reference for the development and utilization of the further research of the armillaria.

1. Introduction

Armillaria mellea is a parthenogenic parasitic fungus of the family Tricholomataceae in the order Hymenomycetes [1], and is also a hemiphyte of the famous Chinese medicinal herbs asparagus and porcupine [2], which is widely distributed in temperate regions of the Northern Hemisphere and is often found on the roots of coniferous or broad-leaved tree trunks in summer and autumn. It has a variety of bioactive components, including polysaccharides, sterols, sphingolipids, fatty acids, sesquiterpenes, non-hallucinogenic indole compounds, peptides, enzymes, adenosine derivatives and more than 110 others, among which polysaccharides are proven to play a key role, A. mellea polysaccharides, which can be extracted and taken out from the mycelium, ascospores, mycelium and fermentation broth of A. mellea [3-6]. With the development of biotechnology and in-depth research on the biological activity of A. mellea polysaccharides, more and more studies have shown that A. mellea polysaccharides have a wide range of biological activity and application value. Therefore, a comprehensive introduction and discussion of the physicochemical properties, extraction methods and their biological activities of A. mellea polysaccharides is of great significance to promote the in-depth research and application of A. mellea polysaccharides.

2. Extraction Process and Optimization of Polysaccharide from A. Mellea

Polysaccharide extraction can use different extraction methods according to the characteristics of raw materials and extraction sites and other factors. A variety of methods have been developed to

extract polysaccharide components, mainly including acid-base extraction, hot water extraction, ultrasound-assisted extraction, compound enzyme-assisted extraction, microwave-assisted extraction, ultrasound-microwave-assisted extraction, pressurized extraction, supercritical extraction, etc. [7,8].

Acid-base extraction method, for some polysaccharides with low water solubility, the extraction rate of hot water extraction method is low, and the extraction rate can be increased by adding acid or alkali solution. The acid-base extraction method is influenced by the pH, temperature and extraction time of the solution, and the solution is too acidic or too alkaline to degrade the polysaccharides. Sun Yujiao [9] investigated different extraction methods to extract porcupine tea polysaccharides and found that the alkali extraction of porcupine tea polysaccharides yielded higher than water and acid extraction, and the highest glyoxalate content of porcupine tea polysaccharides was obtained by the alkali extraction method, which was more favorable to extract polysaccharides rich in glyoxalate; Sun Wei Xuan [10] used acid, alkali and enzymatic methods to extract potato pomace pectin polysaccharides and found that the alkali extraction method yielded the highest polysaccharides (23.1%).

Hot water extraction method, i.e. water extraction and alcoholic sedimentation method, is the traditional method of polysaccharide extraction, the required reagents and equipment are simple and easy to obtain, easy to operate, can avoid the destruction of polysaccharide by acid extraction method and alkali extraction method, and the polysaccharide activity of honeysuckle extracted by this method is high. The raw material is destroyed by the continuous action of hot water, which makes the intracellular polysaccharide flow out. The extraction rate of polysaccharide was affected by temperature, material-liquid ratio and extraction time. Du Guofeng [11] et al. used hot water extraction method to extract *Astragalus* polysaccharides, and compared the temperature, extraction time, and feed-liquid ratio, and found that the polysaccharide yield was proportional to temperature, extraction time, and feed-liquid ratio within a certain range. The optimum extraction conditions were 90 °C, 2.5 h extraction time, and 1:8 g/mL ratio; when the conditions continued to increase, the polysaccharide yield decreased; By investigating the effect of different methods on the extraction rate of polysaccharides from *A.mellea*, Lu Qi [12] found that hot water extraction method gave higher extraction rate than ultrasonic extraction and homogeneous extraction, and the polysaccharide yield was less affected by the particle size of raw materials.

Ultrasound-assisted extraction method is to destroy the cell wall structure by high-frequency vibration of ultrasound in the liquid to promote the penetration of solvent and the release and diffusion of active ingredients, which has the advantages of simpler operation, less reagent dosage, less time consuming, lower temperature and significantly higher extraction efficiency compared with the hot water extraction method, but requires higher equipment. Ultrasound can effectively destroy the cell wall, so that the particle size of the extract can be reduced, the transfer of the cell contents can be enhanced, and the extraction rate of honeysuckle polysaccharide increases [13]. Wang Jiachu [14] et al. used ultrasound-assisted hot water extraction process to extract polysaccharides from *A.mellea*, and promoted the rapid dissolution of polysaccharides by ultrasound on the basis of simple and environmentally friendly operation. Although this method can enhance the polysaccharide extraction rate, it suffers from the problem that the heat production effect is not strong, which easily leads to insufficient extraction temperature.

Enzymatic extraction is a method to extract intracellular polysaccharides by disrupting the cell wall structure of raw materials under the action of enzymes, which can save time and obtain stable products. Enzymatic extraction is usually carried out in synergy with ultrasound, microwave or using complex enzymes, which can further enhance the extraction efficiency. With the synergistic effect of complex enzymes, the hydrolysis of raw materials is more complete and the polysaccharide extraction rate is improved. The mild reaction of compound enzyme method can

better preserve the polysaccharide activity, which is a mild and efficient extraction method, but the enzyme price is high, and the extraction cost is large and affected by the enzyme addition, enzyme ratio, pH, temperature, etc. The optimal extraction conditions for the extraction of crude polysaccharides from *M. gaultheriae* using a complex enzyme method by Peiwen Su [15] were: particle size 80 mesh, enzyme addition (mass ratio) 4%, extraction time 2 h, pH=5, temperature 45 °C, enzyme ratio (papain: cellulase: pectinase) 2:2:1, material-liquid ratio 1:50 g/mL, and crude polysaccharide extraction rate of $18.86 \pm 0.04\%$.

Microwave-assisted extraction is the extraction of polysaccharides by microwave thermal effect, the principle is to use the combination of microwave and traditional hot water extraction method. The microwave power and extraction time will affect the polysaccharide extraction rate. Microwave power, extraction time, etc. will affect the polysaccharide extraction rate, polysaccharide yield and microwave power and extraction time are positively correlated, but too much power and too long time tend to make polysaccharide degradation. Liyuan Feng [16] extracted pectin polysaccharides from broccoli stems by microwave-assisted acid method with pH and microwave power as highly significant factors, and the polysaccharide yield was 4.8%. Jiaolong Fu [17] used microwave-assisted extraction and hot water extraction to extract pumpkin polysaccharides and found that the extraction rate of microwave-assisted extraction method was 5.25 % higher than that of hot water extraction method.

Ultrasonic-microwave-assisted extraction is the extraction of polysaccharides by using ultrasonic cavitation effect and microwave thermal effect synergistically, which can improve the extraction rate. Supercritical extraction method is an extraction method that uses supercritical fluid with similar penetration to gas and similar solubility to liquid for extraction and separation, which is suitable for separating heat-sensitive substances. Although this method has high extraction efficiency, the equipment is complicated and costly, so it cannot be widely used. The pressurized solvent method can accelerate the extraction of polysaccharide components by heating up the solvent under pressure and strengthening the penetration and diffusion ability of the solvent to the cells [18]. This method can accelerate the extraction of polysaccharide components.

The response surface analysis method can determine the optimal process conditions for the extraction of polysaccharides from *A.mellea*, thus enhancing the yield of polysaccharides obtained. It was found that the optimal conditions of each extraction method could be analyzed and screened by using factors such as extraction temperature, extraction time and liquid to material ratio as experimental factors and the extraction rate of honeysuckle polysaccharide as response values. Different extraction methods can also add experimental factors to participate in the analysis appropriately, such as the ultrasonic power for the ultrasound method, the amount of enzyme added for the enzyme method, and the pressure for the pressurization method.

3. Structure and Physicochemical Properties of *A.mellea* Polysaccharide

3.1. Composition Structure of *A.mellea* Polysaccharide

Polysaccharide is the main active component of *A.mellea*, which mainly includes intracellular polysaccharide and extracellular polysaccharide. The polysaccharide content of *A.mellea* varies in different developmental stages, and the experimental study showed that the polysaccharide content of *A.mellea* was 9% in mycelium, 2.27% in zygote, 1.12% in corpuscle and 0.87 g/100 mL in fermentation broth, indicating that the polysaccharide content of mycelium was the highest [19]. The fermentation broth contained 0.87 g/100 mL, indicating that the mycelium contained the most polysaccharides. The composition of the polysaccharides obtained from the fermentation broth also differed greatly depending on the extraction method of *A.mellea*.

Due to the different cultivation methods and extraction processes of *A.mellea*, the results

obtained by different scholars are not the same. Hong Yi [20] et al. isolated the intracellular polysaccharide of *A.mellea* from its mycelium, and the polysaccharide fraction reacted pink with carbazole ethanol (Dische reaction), and the analysis proved that it contained glyoxalate, and the constituent monosaccharides were D-glucose, D-galactose, D-mannose, and D-xylose by paper chromatography and gas chromatography, and the constituent monomers were pyranose and did not contain protein. Chen-Chen Yu [21] et al. analyzed the structure of water-soluble polysaccharides in the fermentation broth of *A.mellea* using UV and IR spectroscopy and gas phase derivatization, and the results showed that the water-soluble polysaccharides were single in composition and all of them were pyranose type glucose. Xiaojie Liu [22] et al. obtained three homogeneous polysaccharides, AMFP-I, AMFP-II and AMFP-III, which were mainly composed of galacturonic acid, glucose, galactose and xylose, from the fermentation broth of *A.mellea* by dextran gel chromatography.

3.2. Physicochemical Properties of *A.mellea* Polysaccharide

Since the processes of extraction methods vary, there may be compositional differences in the *A.mellea* polysaccharides extracted under different conditions, so it is extremely necessary to analyze the physicochemical properties of *A.mellea* polysaccharides. The molecular mass, monosaccharide composition and glycosidic bond type of polysaccharides can be analyzed by hydrolysis, infrared spectroscopy, mass spectrometry, high performance liquid chromatography and gas chromatography in general. The analysis revealed that the chemical composition of *Nectarobacterium* polysaccharides is diverse in structure and has different biological activities. Two polysaccharide fractions with α -glucan structure were isolated from the substrates of *A.mellea*, which could reduce cholesterol and triglyceride concentrations, thus reducing the risk of cardiovascular diseases; the polysaccharide isolated from the sporophore of *A.mellea* had good antioxidant ability and its carbohydrate content was more than 58%; the polysaccharide isolated from the (1 \rightarrow 6)-linked- α -d-glucan was more than 58%.) -linked- α -d-glucopyranosyl and (1 \rightarrow 2,6)-linked- α -d, another *A.mellea* polysaccharide with immunomodulatory function [glycosyl and (1 \rightarrow 2,6)-linked- α -d [23].

4. Biological Activity of *A.mellea* Polysaccharide

4.1. Antioxidant Activity

The significant antioxidant activity of *A.mellea* polysaccharide is determined by its special chemical structure and molecular weight. Its ability mainly comes from the various active groups contained in polysaccharide molecules, such as hydroxyl, methyl and aldehyde groups, which can react with free radicals, reduce their production, and thus play a protective role against free radical damage to cells and tissues. The results of traditional chemical antioxidant experiments on *M. gaulis* showed that Ag-1 and Ag-2 both have the ability to scavenge DPPH radicals, O₂--radicals, ABTS radicals and reduce Fe³⁺, and it was found that both Ag-1 and Ag-2 have protective effects on HepG2, and Ag-1 is more effective, which proves that it has antioxidant activity [15] Fangjuan Zhao [24] et al. performed progressive high temperature tolerance domestication of three strains of *A.mellea*, and both the domesticated *A.mellea* and parental *A.mellea* showed different degrees of antioxidant capacity. Lianyu Zhou et al. studied the antioxidant activity and antibacterial activity of selenium polysaccharide of *A.mellea* yellow-green [25] showed that the scavenging ability of selenium polysaccharide on hydroxyl radical, DPPH radical and ABTS⁺ radical increased significantly (P<0.05) with the increase of its concentration, but the scavenging rate was less than that of ascorbic acid.

4.2. Antitumor Activity

A.mellea polysaccharide has a significant effect on the treatment of tumors. It can induce apoptosis of cancer cells through various pathways, thus exerting anti-tumor effects. It was found that polysaccharide extracted from A.mellea could inhibit the growth of H22 hepatocellular carcinoma cells and induce apoptosis of hepatocellular carcinoma cells by regulating the expression of Bax, Bcl-2, caspase-3 and other proteins. The polysaccharide from A.mellea could inhibit the proliferation of SMMC-7721 hepatocellular carcinoma cells in vitro, inhibit intracellular protein synthesis and inhibit Bcl-2 protein expression to promote apoptosis [26]. In lung cancer patients, A.mellea polysaccharide can improve SOD, GSH-Px, and GSH activity in the body, increase serum SOD, GSH-Px, and GSH levels in patients, reduce MDA levels, and decrease the damage caused by free radicals, which play an important role in the anti-tumor process [27]. In addition, it was found that A.mellea polysaccharide can also improve the activity of T lymphocytes and the killing effect of NK cells, thus enhancing human immunity and reducing the growth of cancer cells.

4.3. Immunomodulatory Activity

A.mellea polysaccharide has immunomodulatory activity and can act by promoting the phagocytosis of macrophages, enhancing human immunity and regulating immune levels. A.mellea polysaccharide can stimulate the phagocytosis of macrophages, promote the activation of macrophages and secrete inflammatory cytokines. Hanzhen [28] isolated and purified AAMP-A70, a polysaccharide with a molecular weight of 5.6 KDa, from the cotyledons of A. honeysuckle, and showed that AAMP-A70 exerts immunomodulatory activity by activating NF κ B and MAPK signaling pathways through binding to TLR-2 and activating macrophages. In addition, A.mellea polysaccharide can enhance the immune function of the body and improve the activity of T-lymphocytes and B-lymphocytes, thus strengthening the resistance of the body. In a study by Huiguo Wang [29] et al. found that A.mellea polysaccharide enhances the phagocytic function of phagocytes and the proliferation capacity of splenic T lymphocytes, and is an immunomodulator that can improve the immunity of the body. In addition, it has been suggested that A.mellea polysaccharide can enhance immune function by modulating the action of immune cells and immune molecules [30].

4.4. Anti-inflammatory Activity

The anti-inflammatory activity of A.mellea polysaccharides is one of the hot topics of research in recent years. Anti-inflammatory effect refers to the inhibition of cellular and molecular activities in the inflammatory process, thus reducing the inflammatory response and tissue damage. Several studies have shown that A.mellea polysaccharides are able to exert their anti-inflammatory effects by inhibiting the activity of inflammatory cells, modulating the immune system and scavenging free radicals in various ways [6]. A.mellea polysaccharide reduced the inflammatory response of periodontitis in mice by regulating the gene expression of pro-inflammatory factors IL-1 β , TNF- α and anti-inflammatory factor IL-10 in periodontal tissues [31]. In laboratory studies, the anti-inflammatory effects of A.mellea polysaccharides have been initially demonstrated. Through in vivo and in vitro experiments, researchers found that A.mellea polysaccharide was able to reduce the inflammatory response in the liver of mice and lower the level of inflammatory indicators, as well as enhance the function of the immune system of mice. The anti-inflammatory effect of A.mellea polysaccharide is exerted by the synergistic effect of its multiple bioactive components [32]. However, further clinical studies are needed to evaluate the mechanism of action and the

prospect of clinical application.

4.5. Modulation of Glycolipid Metabolic Activity

A.mellea polysaccharide can regulate glucolipid metabolism. A.mellea polysaccharide can improve insulin resistance through the regulation of lipid metabolism. The alkali-extracted polysaccharide AAMP significantly reduced fasting blood glucose levels and improved glucose intolerance in type 2 diabetic rats, and increased the expression levels of lipoprotein lipase (LPL), adipose triglyceride lipase (ATGL) and hormone-sensitive lipase (HSL) by regulating the transcription factor SREBP-1c, and decreased the expression levels of serum triglycerides (TG) and free fatty acids (FFA). Expression levels, thereby inhibiting adipogenesis [31]. In addition, it was shown that the total sugar AMP and neutral sugar AMP-N of A.mellea had significant oral hypoglycemic activity, and the hypoglycemic effect of AMP-N was superior to that of AMP. The mechanism of hypoglycemic effect of AMP-N, on the one hand, increased the level of autophagy in the liver of diabetic mice, reduced fat accumulation, and improved insulin sensitivity; on the other hand, protected the pancreatic islets of diabetic mice and improved the damaged function [33]. On the other hand, it protected the islets of diabetic mice and improved the damaged function.

4.6. Other Roles

In addition to the above effects, many studies have confirmed that A.mellea polysaccharides have biological activities such as anti-Alzheimer's disease [34], anti-aging, neuroprotective, hepatoprotective, and antibacterial [35].

5. Summary and Outlook

A.mellea is one of the famous traditional Chinese herbal medicines for tonicity and disease prevention in China, which has a long history of application in tonicity and disease prevention. In recent years, the biological activity of fungal polysaccharides has attracted more and more attention, and the extraction, structure identification, pharmacological activity and mechanism of action of A.mellea polysaccharides have also become hot spots for research. At present, there are many experimental studies on the pharmacological effects of A.mellea polysaccharides, but most of them are on the whole animal level, and the studies on the mechanism of action at the cellular and molecular levels still need to be explored in depth.

In recent years, a lot of research has been done at home and abroad on the polysaccharide of A.mellea and its medicinal value, and good research results have been achieved, and nutritional functional health food such as A.mellea tablets, A.mellea punch and A.mellea syrup with health functions have been developed. More in-depth research and development on A.mellea has been going on, and as the research on A.mellea polysaccharide continues, more preparations will definitely be developed to better serve human health.

Acknowledgment

This work was supported in part by a grant from Jilin Province 2022 College Student Innovation and Entrepreneurship Training Program Project (S202210199056).

References

[1] Chen Zhouli, Wu Xianjin, Tian Yuqiao, Zou Juan & Fang Wei. (2019). Research progress on active ingredients and product development of *Armillaria mellea*. *Modern Food* (21), 25-29.

- [2] Yu Min & Shen Yeshou. (2002). Comparison of polysaccharide content and immunological activity of wild and artificially cultured *Armillaria mellea mycorrhizae*. *Journal of Anhui University (Natural Science Edition)* (01), 107-110.
- [3] Chen Liangwen, Zhang Yunxia, Yu Min & Wang Shunchang. (2013). Study on the mechanism of delaying senescence of *Cryptobacterium hidradenum* by mycorrhizal polysaccharide. *Chinese Herbal Medicine* (04), 449-453.
- [4] Li Shasha, Wang Weifeng, Li Fan & Li Fang (2020). Research progress of Chinese medicine in the treatment of periodontitis. *Northwest Journal of Pharmacy* (02), 313-317.
- [5] Cheng Ran, Wu Zhiwu, Li Mingming, Shao Meiyong & Hu Tao (2020). Interleukin-1 β is a potential therapeutic target for periodontitis: a narrative review. *International Journal of Oral Science* (01), 1-9.
- [6] Shen Shoudong & Shen Minghua. (2018). Effect of *Armillaria mellea* polysaccharide on inflammatory response and apoptosis in arterial thrombosis rats. *Journal of Medicine, Yanbian University* (01), 4-7.
- [7] Liu Yinze., Li Peilei, Qiao Liancheng & Wu Hengmei (2021). Progress in the extraction process and bioactivity of red pine polysaccharides. *Chinese Wild Plant Resources* (09), 37-40+64.
- [8] Chen Ruizhan, Ren Xing, Yin Wei & Luo Shujun (2020). Ultrasonic disruption extraction, characterization and bioactivities of polysaccharides from wild *Armillaria mellea*. *International Journal of Biological Macromolecules*.
- [9] Sun Yujiao, Ma Yunhao, Wang Fan, Yuan Xushuang, Xu Yang, Zhang Nan & Wang Jiankang (2021). Effects of different extraction methods on the physicochemical properties and antioxidant effects of porcupine tea polysaccharides. *Journal of Shaanxi University of Science and Technology* (05), 31-38.
- [10] Sun Weixuan, Tian Jinhu, Chen Jianle, Chen Shiguo, Liu Donghong & Ye Xingqian. (2021). Effect of extraction methods on the composition and molecular chain conformation of pectin polysaccharides from potato pomace. *Chinese Journal of Food Science* (07), 216-224.
- [11] Du Guofeng & Jiang Ning. (2020). Study on the extraction of *Astragalus* polysaccharides. *Light Industry Science and Technology* (10), 36-37+47.
- [12] Lu Qi, Xue Shujing, Yang D, Wang Shaohua & Li Lu. (2021). Effect of three extraction methods on the antioxidant properties of crude polysaccharides from *Agaricus grandis*. *Food Science and Technology* (11), 171-178.
- [13] Nuerxiati R., Abuduwaili A., Mutailifu P., Wubulikasimu A., Rustamova N., Jingxue C., & Yili A. (2019). Optimization of ultrasonic-assisted extraction, characterization and biological activities of polysaccharides from *Orchis chusua D. Don* (Salep). *International Journal of Biological Macromolecules*, 141, 431-443.
- [14] Wang Jiachu, He Zeliang, Shi Chuang, Zhang Jing, Xu Na & Bao Honghui. (2018). Optimization of ultrasonic-assisted extraction process of water-soluble polysaccharides from hazel mushroom by response surface methodology. *China Food Additives* (07), 65-70.
- [15] Su Peiwen. (2022). Extraction, purification and antioxidant activity of *Armillaria gallicapolsaccharide* (Master's thesis, Jilin University).
- [16] Feng Liyuan, Liu Jinglun, Yang Meifu, Ai Wei & Wang Yujie. (2022). Optimized extraction of pectin polysaccharides from broccoli stems by response surface microwave-assisted acid method. *Food Research and Development* (08), 124-132.
- [17] Fu Jiaolong, Liu Jing, Yang Tianyi, Hu Cuiying & Li Liangzhi (2011). Study on the extraction process of pumpkin polysaccharide. *Food Industry* (12), 38-42.
- [18] Zou Fanghua. (2017). Preparation process of polysaccharide of wild honeysuckle from Heilongjiang and its functional study (Master's thesis, Heilongjiang University).
- [19] Chen Xiaomei, Guo Shunxing, Wang Qiuying, Xiao Peigen. (2001). Study on the polysaccharide composition of Honeysuckle at different developmental stages. *Chinese Journal of Traditional Chinese Medicine* (06).
- [20] Hong Yi, Shen Yeshou, Fan Yeyang. (1998). Isolation and purification of *Mycobacterium* honeysuckle polysaccharide and some of its physicochemical properties. *Chinese Journal of Pharmaceutical Sciences* (09).
- [21] Yu Chenchen, Zhu Xianfeng, Fan Shujun & Qin Renbing. (2011). Isolation and identification of water-soluble polysaccharides from honeysuckle and optimization of fermentation conditions. *Food Science and Technology* (06), 84-87.
- [22] Liu Xiaojie, Jiao Lianqing, Yang Limin, Yu Min & Chen Nan (2012). Isolation and purification of polysaccharides from Nectarine bacteria and HPLC analysis of PMP derivatization. *Chinese Pharmacist* (04), 448-451.
- [23] Zhang Shanshan, Liu Xiaoqian, Yan Lihua, Zhang Qiwei, Zhu Jingjing, Huang Na, & Wang Zhimin. (2015). Chemical compositions and antioxidant activities of polysaccharides from the sporophores and cultured products of *Armillaria mellea*. *Molecules*, 20 (4), 5680-5697.
- [24] Zhao Fangjuan, Deng Wan, Xie Xiuchao & Yang Xueying. (2022). Extracellular polysaccharide content and antioxidant activity of high temperature resistant honeysuckle. *Journal of Shaanxi University of Technology (Natural Science Edition)* (05), 79-85.
- [25] Zhou Lianyu, Zhong Rui, Jiao Lu, Chen Q. H. & Zhan Yan. (2020). Study on antioxidant activity and antibacterial activity of selenium polysaccharide from *Methanobacterium yellowish green*. *Food Research and Development* (15), 6-10.

- [26] Song Chengzhi & Xu Yan. (2010). Study on the chemical composition and pharmacological effects of Honeysuckle. *Anhui Agricultural Science* (10), 5119-5120.
- [27] Wu Jun, Li Junzhe, Huang Congcong, Zhang Pan, Lin Ximing, Cheng Quan & Zhang Guochao. (2019). Effect of honeysuckle polysaccharide on SOD, GSH-Px, GSH and MDA levels in lung cancer patients and analysis of the causes. *PLA Journal of Preventive Medicine* (08), 115-116+119.
- [28] Han Zhen. (2018). Study on the immunomodulatory activity of honeysuckle dextran on RAW264.7 cells (Master's thesis, Northeast Normal University).
- [29] Wang Huiguo & Feng Baomin. (2009). Experimental study on the immunomodulatory activity of polysaccharide from *Methanosarcina* honeysuckle. *Journal of Shaanxi University of Science and Technology (Natural Science Edition)* (02), 62-64.
- [30] Wang H. G., Guan H. Q., Zhao I. N. & Li X. H. (2007). Effect of honeysuckle polysaccharide on serum IL-2 and TGF- β 1 levels in mice. *Advances in Modern Biomedicine* (09), 1306-1307.
- [31] Tian Yue. (2022). Effects of *Armillaria Mellea* Polysaccharide on Inflammation and Alveolar Bone Resorption in Experimental Periodontitis Mice (Master's thesis, Jilin University).
- [32] Geng Yan, Zhu Shuiling, Lu Zhenming, Xu Hongyu, Shi Jinsong, & Xu Zhenghong (2014). Anti-inflammatory activity of mycelial extracts from medicinal mushrooms. *International journal of medicinal mushrooms*, 16 (4), 319–325.
- [33] Yang Siwen, Meng Yuhan, Yan Jingmin, Wang Na, Xue Zhujun, Zhang Hang, & Fan Yuying. (2018). Polysaccharide-enriched fraction from *Amillariella mellea* fruiting body improves insulin resistance. *Molecules*, 24 (1), 46.
- [34] Zhang Zhiyuan, Wang Shuai, Tan Haining, Yang Pei, Li Yuanyuan, Xu Lingchuan & Liu Yuhong. (2022). Advances in polysaccharides of natural source of the anti-Alzheimer's disease effect and mechanism. *Carbohydrate Polymers*, 119961.
- [35] Li Hongyu, Xu Guangyu & Yuan Guangxin. (2022). Effects of an *Armillaria mellea* Polysaccharide on Learning and Memory of D-Galactose-Induced Aging Mice. *Frontiers in Pharmacology*.